

JC595 U.S. PTO
08/31/99

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE
REQUEST FOR FILING CONTINUATION/DIVISIONAL
APPLICATION UNDER 37 C.F.R. § 1.53(b)

JC675 U.S. PTO
09/30/99

Box PATENT APPLICATION
Assistant Commissioner for Patents
Washington, D.C. 20231

Sir:

This is a request for filing a [] continuation [X] divisional application under 37 C.F.R.
§ 1.53(b) of pending Application No. 08/476,120 filed on June 7, 1995, for PEPTIDES AND
POLYPEPTIDES DERIVED FROM THE SUBMAXILLARY GLAND OF THE RAT.
CORRESPONDING POLYCLONAL AND MONOCLONAL ANTIBODIES, CORRESPONDING
HYBRIDOMAS AND USES OF THESE PRODUCTS FOR DIAGNOSIS, FOR DETECTION OR
THERAPEUTIC PURPOSES, by the following named inventor(s):

- (a) Full Name Isabelle ROSINSKI-CHUPIN
- (b) Full Name Diana TRONIK
- (c) Full Name Francois ROUGEON
- (d) Full Name Nabil SEIDAH

[X] The entire disclosure of the prior application from which a copy of the oath or declaration is
supplied herewith is considered as being part of the disclosure of the accompanying
application and is hereby incorporated by reference therein.

[] This application is being filed by less than all the inventors named in the prior application.
In accordance with 37 C.F.R. 1.63(d)(2), the Commissioner is requested to delete the
name(s) of the following person or persons who are not inventors of the invention being
claimed in this application.

- (a) Full Name _____
- (b) Full Name _____
- (c) Full Name _____

- 1. [X] Enclosed is a copy of the prior Application No. 08/476,120 as originally filed on
June 7, 1995, including copies of the specification, claims, drawings and the
executed oath or declaration as filed.

2. ☐ Enclosed is a revised prior application and a copy of the prior executed oath or declaration as filed. No new matter has been added to the revised application.
3. ☐ _____ statement(s) claiming small entity status ☐ are enclosed ☐ were filed in prior Application No. __, filed on __.
4. ☒ The filing fee is calculated below ☒ and in accordance with the enclosed preliminary amendment:

CLAIMS					
	NO. OF CLAIMS		EXTRA CLAIMS	RATE	FEE
Basic Application Fee					\$760.00
Total Claims	3	MINUS 20 =	-0-	x \$18.00 =	-0-
Independent Claims	2	MINUS 3 =	-0-	x \$78.00 =	-0-
If multiple dependent claims are presented, add \$260.00					-0-
Total Application Fee					\$760.00
If small entity status is claimed, subtract 50% of Total Application Fee					-0-
Add Assignment Recording Fee of \$40.00 if Assignment document is enclosed					-0-
TOTAL APPLICATION FEE DUE					\$760.00

5. ☐ Charge \$ _____ to Deposit Account No. 02-4800 for the fee due.
6. ☒ A check in the amount of \$ 760.00 is enclosed for the fee due.
7. ☒ The Commissioner is hereby authorized to charge any appropriate fees under 37 C.F.R. §§ 1.16, 1.17 and 1.21 that may be required by this paper, and to credit any overpayment, to Deposit Account No. 02-4800. This paper is submitted in triplicate.
8. ☒ Cancel in this application original claims 2-14 of the prior application before calculating the filing fee. (At least one original independent claim must be retained for filing purposes.)

9. ☐ Amend the specification by inserting before the first line the sentence: --This application is a ☐ continuation, ☐ divisional, of Application No. _____, filed _____. --
10. ☐ Transfer the drawings from the pending prior application to this application and abandon said prior application as of the filing date accorded this application. A duplicate of this paper is enclosed for filing in the prior application file. (May only be used if signed by person authorized under 37 C.F.R. § 1.138 and before payment of issue fee.)
11. ☐ New drawings are enclosed.
12. ☒ Priority of Application No. 88/13353 filed on October 11, 1988 in France (country), and International Application No. PCT/FR88/00523, filed on October 1, 1989 are claimed under 35 U.S.C. § 119.
- ☐ The certified copy of the priority application
☐ is enclosed
☐ was filed on ____ in prior Application No. __, filed on ____
☐ has not yet been filed.
13. ☒ A preliminary amendment is enclosed.
14. ☒ Also enclosed are an Information Disclosure Statement Transmittal Letter; an Information Disclosure Statement; and a PTO Form 1449 listing 24 documents.
15. ☒ The power of attorney in the prior application is to Burns, Doane, Swecker & Mathis, L.L.P..
- a. ☒ The power appears in the original papers in the prior application.
- b. ☐ Since the power does not appear in the original papers, a copy of the power in the prior application is enclosed.
- c. ☒ Recognize as Associate Attorney Susan M. Dadio, Reg. No. 40,373.

- d. ☒ Address all future communications to: (May only be completed by applicant, or attorney or agent of record.)

Regis E. Slutter
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August 31, 1999
Date

By: Regis E. Slutter
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☐ assignee of complete interest
☒ attorney or agent of record
☐ filed under 37 C.F.R. § 1.34(a)

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Patent Application of)
)
Isabelle ROSINSKI-CHUPIN et al.) Group Art Unit: Unassigned
)
Application No.: Divisional of Application)
Serial No.: 08/476,120) Examiner: Unassigned
)
Filed: August 31, 1999)
)
For: PEPTIDES AND POLYPEPTIDES)
DERIVED FROM THE SUBMAX-)
ILLARY GLAND OF THE RAT. . .)
FOR DIAGNOSIS, FOR DETECTION)
OR THERAPEUTIC PURPOSES)

PRELIMINARY AMENDMENT

Assistant Commissioner for Patents
Washington, D.C. 20231

Sir:

Prior to the examination on the merits, please amend the above-identified
application as follows:

IN THE SPECIFICATION:

Please insert Figure 1 enclosed herewith on a separate sheet.

On page 1, under the title and before line 6, please insert the following:

--CROSS-REFERENCE TO RELATED APPLICATIONS

This application is a divisional of application Serial No. 08/476,120, filed on June 7, 1995, which is a continuation of application Serial No. 08/153,277, filed November 17, 1993, now United States Patent No. 5,859,189, which is a continuation of application Serial No. 07/499,276, filed July 19, 1990, now abandoned.

BACKGROUND OF THE INVENTION--.

On page 1, in between lines 22 and 23, kindly insert the following heading:

--SUMMARY OF THE INVENTION--.

On page 1, in between lines 31 and 32, kindly insert the following:

--BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1A shows mRNAs coding for SMR1 in the SMG of male rats, female rats treated with testosterone, castrated male rats, castrated male rats treated with testosterone and female rats.

Figure 1B shows mRNAs coding for SMR1 from various amounts of RNA from the SMG of male rats and female rats.

DETAILED DESCRIPTION OF THE INVENTION--.

On page 9, line 21, please delete the first seven amino acids which are illegible and insert the following: --Thr Lys Thr Asp Ala Lys Ile--.

IN THE CLAIMS:

Please cancel claim 1, without prejudice or disclaimer to the subject matter disclosed therein.

Please add the following new claims 15-17 as follows:

--15. A method for detecting and/or quantifying a nucleic acid coding for SMR1 comprising:

- a) contacting a DNA or RNA molecule with a nucleotide probe having the nucleotide sequence of SEQ ID No. 1 under conditions allowing said probe to hybridize with the target DNA or RNA molecule; and
- b) detecting the hybrid formed between the DNA or RNA molecule and said nucleotide probe.

16. A method according to claim 15 for detecting and/or qualifying mRNA coding for SMR1 polypeptide consisting essentially of:

- a) preparing mRNAs containing poly (A) sequence from the sub-maxillary gland of a rat;
- b) subjecting said RNAs prepared in step a) to an electrophoresis in a gel;
- c) transferring said RNAs to a nylon membrane;

- d) contacting said membrane with a nucleotide probe having the nucleotide sequence of SEQ ID No. 1 under conditions allowing said probe to hybridize to the target RNA;
- e) detecting the hybrid formed between the RNA and the nucleotide probe; and
- f) optionally quantifying the number of hybrid molecules as formed.

17. A process for testing behavioral characteristics in a patient consisting of assaying SMR1 peptide in a biological fluid of said patient.--

REMARKS

Entry of the foregoing and prompt and favorable consideration of the subject application are respectfully requested.

By the foregoing Amendment, the specification has been amended to include proper headings in accordance with standard United States practice and to update the status of the related applications.

Further, as described in both application Serial Nos. 08/153,277 and 08/476,120, the parent application was a national stage application of PCT/FR89/00523 which designated the United States. The PCT application when filed contained one (1) sheet of drawings. See page 4 of the Request for International Application Under the Patent

Cooperation Treaty (PCT) attached hereto as Exhibit A. This is further evidenced by the fact that the PCT application was published with Figure 1A and B. See WO 90/03981 attached hereto as Exhibit B. On June 11, 1990, applicants filed a request in the United States to begin examination procedures pursuant to 35 U.S.C. § 371 in the parent application.

In view of the above, it is clear that the parent application was originally filed with figures.

Accordingly, by the present amendment, the figures as well as a brief description of the drawings have been inserted into the specification. Since the original application contained such figures, no new matter has been added.

Additionally, new claims 15-17 have been added. Support for these claims can be found throughout the originally filed application. Thus, no new matter has been added.

In the event that there are any questions relating to this Preliminary Amendment, or the application in general, it would be appreciated if the Examiner would telephone the

Application Serial No. Unassigned
Attorney's Docket No. 004900-169

undersigned attorney concerning such questions so that prosecution of this application may
be expedited.

Respectfully submitted,

BURNS, DOANE, SWECKER & MATHIS, L.L.P.

By: Susan M. Dadio
Susan M. Dadio
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Date: August 31, 1999

Peptides and polypeptides derived from the submaxillary gland of the rat, corresponding polyclonal and monoclonal antibodies, corresponding hybridomas and uses of these products for diagnosis, for detection or therapeutic purposes.

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The present invention relates to novel peptides which are maturation products of polypeptides secreted by the submaxillary gland of the rat and analogues of these peptides.

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Many polypeptides having defined biological properties are synthesized in large quantities in the submaxillary gland (SMG) of rodents, and in particular in the SMG of the mouse. These proteins, comprising the nerve growth factor (NGF), the epidermal growth factor (EGF) and renin, have a certain number of properties in common. They are all synthesized in the same type of cell, namely the cells of the convoluted tubules GCT (granular convoluted tubular), in response to various hormonal stimuli, in particular to androgens. Furthermore, it is possible to observe the presence of these secretory proteins in the saliva of the mouse, and they are synthesized in the form of precursors which become active after maturation processes which may involve proteases of the kallikrein type. Some of these proteases of the kallikrein type are also synthesized in the SMG under the control of androgens.

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Attempts at the characterization of the genes controlled by the androgens in the SMG of the rat have led the inventors to analyse the electrophoretic profile of the in vitro translation products of the mRNA of this tissue.

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A specific mRNA of the submaxillary gland of the male rat has been isolated. This mRNA corresponds to a polypeptide which has been designated SMR1. This polypeptide gives maturation products with physiological activity. The present invention relates mainly to these peptides and to other peptides having similar properties.

Thus the subject of the present invention is peptides of formula:

X-His-Asn-Pro-Y

I

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in which:

X represents a glutamine (Gln) or pyroglutamic acid (pyro-Glu) residue,

Y represents a OH group or a residue of a basic amino acid.

The basic amino acids may be lysine or arginine.

More specifically, the subject of the present invention is peptides of formula:

Gln-His-Asn-Pro II

pyro-Glu-His-Asn-Pro III

Gln-His-Asn-Pro-Arg IV

pyro-Glu-His-Asn-Pro-Arg V

Gln-His-Asn-Pro-Lys VI

pyro-Glu-His-Asn-Pro-Lys VII

Another subject of the present invention is the polypeptide SMR1 which gives the maturation products of formulae II, III, IV and V. This polypeptide corresponds to the formula:

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Met Lys Ser Leu Tyr Leu Ile Phe Gly Leu Trp Ile Leu
20
Leu Ala Cys Phe Gln Ser Gly Glu Gly Val Arg Gly Pro
30
Arg Arg Gln His Asn Pro Arg Arg Gln Gln Asp Pro Ser
40
Thr Leu Pro His Tyr Leu Gly Leu Gln Pro Asp Pro Asn
50
60
Gly Gly Gln Ile Gly Val Thr Ile Thr Ile Pro Leu Asn
70
Leu Gln Pro Pro Arg Val Leu Val Asn Leu Pro Gly Phe

80
 Ile Thr Gly Pro Pro Leu Val Val Gln Gly Thr Thr Glu
 90
 100
 Tyr Gln Tyr Gln Trp Gln Leu Thr Ala Pro Asp Pro Thr
 5
 110
 Pro Leu Ser Asn Pro Pro Thr Gln Leu His Ser Thr Glu
 120
 130
 Gln Ala Asn Thr Lys Thr Asp Ala Lys Ile Ser Asn Thr
 10
 140
 Thr Ala Thr Thr Gln Asn Ser Thr Asp Ile Phe Glu Gly
 Gly Gly Lys

15 Another subject of the present invention is monoclonal and polyclonal antibodies directed against the peptides and the polypeptide according to the invention.

20 Another subject of the present invention is hybridomas producing monoclonal antibodies directed against the peptides I to VII and the polypeptide according to the invention.

25 Another subject of the present invention is a procedure for the assay or detection of the peptides and polypeptides according to the invention in tissues and biological fluids which comprises the utilization of monoclonal or polyclonal antibodies according to the invention.

30 For this purpose, it is possible to utilize in particular a method of the RIA type using a peptide labelled by a radioisotope, and the competition between this peptide and the peptide to be assayed (Niswender G.D. et al; Proc. Soc. Exp. Biol. 128, 807, 1968). It is also possible to utilize a method of the ELISA type using, for example, a peptide bound to a support and the competition between this peptide and the peptide to be assayed for antibodies prepared against this peptide. The antibodies retained by the peptide bound to the support are detected by antibodies directed against the former and linked to an enzyme (method derived from Avromeas J. and Guilbert B, C.R. Acad. Sci. Paris 1971, 273, 2305).

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The peptides according to the present invention may be prepared in a standard manner by peptide synthesis in liquid or solid phase by successive couplings of the different amino acid residues to be incorporated (from the N-terminal end toward the C-terminal end in liquid phase, or from the C-terminal end to the N-terminal end in solid phase) and the N-terminal ends and the reactive side chains of which are blocked beforehand by standard groupings.

For this synthesis on solid phase it is possible to utilize in particular the technique described by Merrifield, in the article entitled "Solid phase peptide synthesis" (J. Am. Chem. Soc., 85, 2149-2154).

In order to produce a peptide chain according to the Merrifield procedure, recourse is had to a very porous polymeric resin, to which the first C-terminal amino acid of the chain is bound. This amino acid is bound to the resin through the intermediary of its carboxyl group and its amino function is protected, for example by the t-butoxycarbonyl group.

When the first C-terminal amino acid is thus bound to the resin, the protecting group is removed from the amine function by washing the resin with an acid. In the case in which the protecting group for the amine function is the t-butoxycarbonyl group, it may be removed by treatment of the resin with the aid of trifluoroacetic acid.

The second amino acid, which provides the second residue of the desired sequence, is then coupled to the deprotected amine function of the first C-terminal amino acid bound to the chain. Preferably, the carboxyl function of this second amino acid is activated, for example, by means of dicyclohexylcarbodiimide, and the amine function is protected, for example, by the t-butoxycarbonyl.

In this way, the first part of the desired peptide chain is obtained which contains two amino acids and the terminal amine function of which is protected. As previously, the amine function is deprotected and it is then possible to proceed to the attachment of the third residue under similar conditions to those for the addition of the second C-terminal amino acid.

In this manner, the amino acids which will constitute the

peptide chain are attached one after the other to the amine group, deprotected beforehand each time, of the portion of the peptide chain already formed and which is attached to the resin.

When the whole of the desired peptide chain is formed, the protecting groups are removed from the different amino acids constituting the peptide chain and the peptide is cleaved from the resin, for example, with the aid of hydrogen fluoride.

The peptide thus obtained can be purified, for example by means of column chromatography.

The SMR1 peptide or derivatives of this peptide can also be obtained with the aid of the techniques of genetic engineering; they can also be obtained by purification from biological material by means of techniques of chromatography or precipitation similar to those used, for example, for the purification of human growth hormone starting from the hypophysis.

The monoclonal and polyclonal sera can be prepared according to a standard technique. For this purpose the tetrapeptides or the pentapeptides or multimeric derivatives of these peptides can be coupled to immunogenic agents such as KLH (Keyhole Limpet Hemocyanin), ovalbumin, bovine serumalbumin etc, by a coupling agent such as glutaraldehyde. The SMR1 protein and the derivatives of this protein (peptides derived from this protein, or hybrid proteins containing a part or all of SMR1 linked to another protein such as protein A) can also be injected directly.

The immunizations can be performed in a standard manner, for example, in the rabbit and the mouse by injecting into the animal 100 micrograms, for example, of the coupling product in the presence of Freund's adjuvant 3 to 4 times at intervals of 3 weeks. It is thus possible to obtain polyclonal sera in the rabbit.

The hybridomas and the monoclonal antibodies can be obtained by means of the standard procedures.

The isolation of the mRNA, which corresponds to the SMR1 polypeptide, and the properties of the polypeptide and the maturation products will be described in more detail below.

1) MATERIALS AND METHODS

Animals and hormonal treatments

10 weeks old male and female Wistar rats were obtained from Iffa-Credo. The androgens were removed by castration and 10 days later 35 mg of testosterone (Sterandryl retard, Roussel) were injected by the intraperitoneal route in the cases indicated. In the cases indicated, the same dose of testosterone was administered to female rats. 8 weeks old DBA/2 and "Swiss" mice were obtained from the Pasteur Institute.

Extraction of the RNA and in vitro translation

The RNA was prepared from rat and mouse tissues as described in the literature (Tronik D. et al. 1987, EMBO J. 6, 983-987). The in vitro translation of the RNAs was carried out by using the mRNA-dependent translation system as a lysate in the presence of mRNA-dependent reticulocytes (Pelham H. RB et al., 1976, Eur. J. Biochem. 67, 247-256). The products were analysed by electrophoresis on a denaturing polyacrylamide-NaDodSO₄ gel.

Cloning and characterization of the cDNA coding for SMR1

Poly(A) RNA obtained from SMG of male Wistar rats was used as matrix for the reverse transcriptase, and the double-stranded cDNAs were obtained by means of the DNA synthesis system of Amersham, with the protocol supplied by the manufacturer. The double-stranded cDNA was then inserted into the PstI site of pUC9 by the method of the oligo-d(C) ends (Maniatis, T. et al. (1982) in Molecular Cloning: A Laboratory Manual (Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y.) pp 241-242). The host bacteria (bacterial strain DH 5-1 derived from DH1 and yielding high efficiencies of transformation, see Hanatian DNA cloning vol. 1, p. 111) were transformed and the colonies were screened by hybridization with the probes described below. In brief, the mRNA derived from the SMG of males and females was fractionated in a 5-20% sucrose gradient. The fractions enriched in mRNA of low molecular weight (which were shown by in vitro translation to contain

the mRNA coding for SMR1) were precipitated with ethanol, and they were used as matrix for the reverse transcriptase in the presence of dGTP and dCTP radiolabelled with ^{32}P . About 3000 recombinants were screened on filters in duplicate samples with the radiolabelled cDNA obtained from the SMG of male and female rats. The clones selected demonstrated a strong hybridization with the male cDNA probe but a very weak hybridization with the female probe. The recombinant clones were identified by experiments of inhibition of translation of the mRNAs in a cell-free system by DNA-mRNA hybridization (Paterson et al. (1977) Proc. Natl. Acad. Sci. USA, 74, 4370-4374).

Sequencing of the cDNA coding for SMR1

With the aid of various restriction enzymes, the cDNA coding for SMR1 was cut and the fragments obtained were subcloned in the vector M13 mp9. The sequencing of the DNA was carried out by means of the method of termination with dideoxyribonucleotides (Sanger, F. et al. (1977) Proc. Natl. Acad. Sci. USA, 74, 5463-5467).

2) RESULTS

Analysis of the in vitro translation of mRNA prepared from the SMG of rats and mice.

5 μg of total RNA obtained from the SMG of male and female mice or male and female rats were translated in a cell-free system of reticulocytes in the presence of ^{35}S -methionine. The products of in vitro translation were subjected to electrophoresis on a 12.5% sodium dodecylsulfate-polyacrylamide gel and autoradiographed.

The comparison of the products of the in vitro translation of RNA prepared from the SMG of male and female rats shows the presence of several polypeptides (of apparent molecular weights of 18,000, 19,000, 35,000, 46,000) in larger amounts in the male rats than in the females. These data show the existence of a sexual dimorphism in the SMG of the rat.

Furthermore, the comparison of these products of translation with those obtained with RNAs derived from SMG of mice shows that most of the major polypeptides are different in the two species. In

particular, whereas sex-linked differences could also be observed in the mouse, these latter did not relate to the same polypeptides as in the rat. Conversely, the products of translation observed in the rat to be specific for the male seem to be absent in the mouse.

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Isolation and sequence of a cDNA complementary to a specific mRNA of the male rat

In order to isolate mRNAs specific for the male from the SMG of the rat, a bank of cDNA prepared from this tissue was constructed in pUC9. The recombinant clones were screened by utilizing a differential screening strategy such as that described in the Materials and Methods section. The positive clones were characterized by experiments of inhibition of translation of the mRNAs in a cell system by DNA-mRNA hybridization. One category of recombinant cDNAs suppressed the in vitro synthesis of a polypeptide having an apparent molecular weight of 19,000 daltons. This polypeptide, designated SMR1, is present in the products of in vitro translation of RNAs from male rats but is not present in those from females. The corresponding cDNA was utilized as probe in RNA hybridization experiments with transfer to a nitrocellulose membrane. For the hybridization a solution was used containing:

- 0.5 M sodium phosphate, pH 7.2
- 7% sodium dodecylsulfate (SDS)
- 1 mM EDTA
- 1% bovine serum albumin
- sonicated salmon sperm DNA: 100 mg/ml

The experiment was done at 65°C for 16 to 20 h.

4 washings of 10 minutes each were carried out at 65°C with a solution containing:

- 40 mM sodium phosphate, pH 7.2
- 1% SDS
- 1 mM EDTA

The cDNA is hybridized with a mRNA 700 nucleotides long, present in large amounts in the SMG of male rats. This inserted element of cDNA has been characterized in more detail.

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The sequence of the cDNA coding for SMR1 and the sequence deduced for the protein are represented below

5	AAA CTC ACT CAC CAG AGA CCT TCT CAC CAG CAC ATT TCC CCG CTC AGA AGT TTC	54
	<u>Met Lys Ser Leu Tyr Leu Ile Phe Gly Leu Trp Ile</u>	
	TCC AAG CCG CTA CCA AAG ATG AAG TCA CTG TAT TTT ATC TTT GCG CTG TCG ATC	108
	<u>Leu Leu Ala Cys Phe Gln Ser Gly Glu Gly Val Arg Gly Pro Arg Arg Gln His</u>	
	CTT CTA CCA TGC TTC CAG TCA CGT CAG GGT CTC AGA GCG CCA ACA AGA CAA CAT	162
10	<u>Asn Pro Arg Arg</u> Gln Gln Asp Pro Ser Thr Leu Pro His Tyr Leu Gly Leu Gln	
	AAT GGT ACA AGA CAA CAA GAT CCT TCA ACT CTT CCT CAT TAT CTT GGT CTT CAC	216
	<u>Pro Asp Pro Asn Gly Gly Gln Ile Gly Val Thr Ile Thr Ile Pro Leu Asn Leu</u>	
	CCT CAT CCC AAT GGT CCA CAA ATA CCA GTA ACA ATC ACT ATA CCC TTA AAT CTT	270
15	<u>Gln Pro Pro Arg Val Leu Val Asn Leu Pro Gly Phe Ile Thr Gly Pro Pro Leu</u>	
	CAA CCA CCT CGT GTT CTT GTT AAT CTT CCG CGT TTT ATC ACT CCA CCA CCA TTG	324
	<u>Val Val Gln Gly Thr Thr Glu Tyr Gln Tyr Gln Trp Gln Leu Thr Ala Pro Asp</u>	
	CTT GTA CAA CGT ACC ACT GAA TAT CAA TAT CAG TCG CAG CTA ACT GCT CCA CAC	378
20	<u>Pro Thr Pro Leu Ser Asn Pro Pro Thr Gln Leu His Ser Thr Glu Gln Ala Asn</u>	
	CCT ACA CCT CTA AGC AAT CCT CCT ACT CAA CTT CAT TCC ACA GAA CAA CCA AAT	432
	<u>Thr Lys Thr Asp Ala Lys Ile Ser Asn Thr Thr Ala Thr Thr Gln Asn Ser Thr</u>	
	ACA AAA ACA CAT CCC AAA ATC TCC AAC ACT ACT CCG ACT ACC CAA AAT TCC ACT	486
	<u>Asp Ile Phe Glu Gly Gly Gly Lys</u>	
	CAT ATT TTT GAA GGT GGT CCG AAA TAATAAATTCCTTTTCCCACTTACAATACCATAAATCA	549
25	AACACTGTCTACTTTTCCCGCAATAATCTTTAAGCCTTCAGAAACAACCTTTACCCCATATACAAA	620
	TCACAATAAAGACCTAAGCAGCATTACACAGCAAAAA	658

The inserted cDNA has a length of 652 nucleotides, if the poly(A) fragment is excluded. This sequence comprises an open reading frame of 510 nucleotides. The only ATG which can serve as initiation codon is situated 73 nucleotides downstream from the 5' end of the clone of cDNA. The untranslated region at the 3' end (142 nucleotides long) contains the consensus signal for polyadenylation (AATAAA), 23 nucleotides upstream from the poly(A) tail (nucleotides 625-630).

The translation stop codon is found in another AATAAA sequence (512-517) which apparently is not recognized as polyadenylation signal.

The corresponding protein has a length of 146 amino acids, which corresponds to a molecular weight of about 16,000 daltons. This molecular weight is slightly lower than that determined from the electrophoretic mobility of the in vitro translation product (19,000 daltons). Such a difference between the molecular weight calculated from the analysis of the sequence and that determined from the electrophoretic mobility has already been described for other proteins of the SMG of the rat or the mouse.

SMR1 has a relatively high content of glutamine and proline residues, but does not contain a repetitive region. Hence, it does not belong to the family of polypeptides "rich in proline" or "rich in glutamine", which are essential proteins of the SMG. Furthermore, the sequence of the mRNA does not contain at its 5' end the sequence of 80 nucleotides which is characteristic of this family. However, like these proteins, SMR1 does not contain cysteine or methionine residues (except in its signal peptide).

The amino-terminal part of SMR1 is strongly hydrophobic, which is characteristic of signal peptides of most of the secreted proteins. Although the amino-terminal sequence of the mature protein has not been determined directly, from the statistical analysis made according to G. Von Heijne (Nucleic Acids Res. 14, 4683-4690, 1986) (rule "-3,-1"), it may be supposed that the cleavage site of the signal peptide is located between the residues 18 and 19.

The protein SMR1 also shows certain features characteristic of the glycoproteins. The presence of two potential glycosylation sites linked to N are observed at the positions 139 and 136. The protein is relatively rich in proline (12%), threonine (12%) and glutamine (9.5%). Several glycosylation sites linked to O might thus be present in the carboxy-terminal fragment of SMR1, since regions rich in proline and threonine residues are usually present in highly O-glycosylated proteins, such as the mucoproteins and the sialoglycoproteins.

An interesting characteristic is the presence of pairs of

basic amino acids Arg-Arg at positions 27-28 and 33-34. Such dipeptides represent potential sites of cleavage by maturation enzymes (Lazure, C., et al (1983) Can. J. Biochem., Cell Biol. 61, 501-515 and Docherty, K. et al (1982) Ann. Rev. Physiol. 44, 625-638). They flank a tetrapeptide Gln-His-Asn-Pro. The tetrapeptide and its adjacent sequences are located in a hydrophilic environment which renders this region accessible to possible maturation enzymes.

The cleavage of Arg-Arg linkages by a maturation enzyme followed by the removal of the basic residues by carboxypeptidase E (Fricker, L.D. et al (1983) J. Biol. Chem. 258, 10950-10955) and possibly an aminopeptidase (Loh, Y.P. et al (1984) Ann. Rev. Neurosci. 7, 189-222) would produce a mixture of tetrapeptide (Gln-His-Asn-Pro) and pentapeptide (Gln-His-Asn-Pro-Arg), since "Pro-Arg" is not a good substrate for carboxypeptidase E. Other post-translational modifications could also include the formation of pyroGlu acid derivatives of these products, giving rise to a mixture of pyroGlu-His-Asn-Pro-Arg and pyroGlu-His-Asn-Pro. These structures recall those of thyroliberin (TRH).

Regulation of the accumulation of mRNA coding for SMR1 in the SMG of the rat by androgens.

In order to study the regulation of the accumulation of mRNA coding for SMR1 in the SMG of the rat by androgens, mRNA containing a poly(A) sequence was prepared from the SMG of adult males, males castrated 20 days previously, castrated males subjected to a treatment with androgens, females and females treated with androgens. 1 µg of total RNA from male rats, female rats treated with testosterone, castrated male rats, castrated male rats treated with testosterone and female rats was subjected to electrophoresis in a 1.4% agarose-formaldehyde gel, these RNAs were transferred to a Nylon membrane and were hybridized with the cDNA probe coding for SMR1. The time of exposure for autoradiography was 2 hours. The results of the RNA analysis, by transfer of these mRNAs to solid supports, by means of a SMR1 probe labelled with ³²P are shown in the figure (part A). A considerable difference in the accumulation of mRNA coding

for SMR1 is observed in the SMG of male rats and those of females.

Furthermore, various amounts (as indicated in the figure) of RNA from the SMG of male rats and female rats were subjected to electrophoresis on a 2% agarose-formaldehyde gel, they were transferred to filters and hybridized with the cDNA probe coding for SMR1. The film was exposed for 30 hours. As is apparent in part B, the mRNA coding for SMR1 accumulates in very large quantities in the SMG of male Wistar rats, since an amount as low as 1.5 ng of total RNA was sufficient to give a hybridization signal. Conversely, the level of accumulation of mRNA coding for SMR1 in the SMG of female Wistar rats was about 1,000 to 3,000 times lower than that in males.

In the castrated males, the quantity of mRNA coding for SMR1 was reduced 10 to 20 fold. The administration of testosterone to these males restored the amount of mRNA coding for SMR1 to the same value as that observed in the non-castrated males. Furthermore, the administration of testosterone to adult female rats caused the accumulation of mRNA coding for SMR1 in amounts similar to that observed in the males.

A remarkable property of the mRNA coding for SMR1 is its high degree of accumulation in the SMG of the rat in response to a treatment with androgens. This strongly suggests that SMR1 is synthesized in the GCT cells of the SMG (just like the EGF, NGF and renin as well as other proteins under the control of androgens in the SMG of the mouse). Furthermore, the difference in the level of accumulation of mRNA coding for SMR1 in the male and in the female is very great (greater than three orders of magnitude), in comparison with that usually observed for other genes controlled by antigens in the target organs (kidney, liver, SMG).

These results, and in particular the high degree of induction of the SMR1 gene by androgens, suggest that SMR1 may fulfil an important function specific for the male in the rat. SMR1 might be the precursor of a molecule (the tetra- or pentapeptides or the C-terminal part of SMR1) controlling behavioural characteristics in the male rat.

The products described in the present invention can be used for therapeutic purposes or as laboratory reagents.

13

CLAIMS

1. Peptide of formula:

X-His-Asn-Pro-Y

I

in which:

X represents a Gln or pyro-Glu residue,

Y represents a OH group or a residue of a basic amino acid.

2. Peptide according to Claim 1, in which Y is a Lys or Arg residue.

3. Peptide of formula:

Gln-His-Asn-Pro

II

4. Peptide of formula:

pyro-Glu-His-Asn-Pro

III

5. Peptide of formula:

Gln-His-Asn-Pro-Arg

IV

6. Peptide of formula:

pyro-Glu-His-Asn-Pro-Arg

V

7. Peptide of formula:

Gln-His-Asn-Pro-Lys

VI

8. Peptide of formula:

pyro-Glu-His-Asn-Pro-Lys

VII

9. Polypeptide of formula:

1
Met Lys Ser Leu Tyr Leu Ile Phe Gly Leu Trp Ile Leu

20

Leu Ala Cys Phe Gln Ser Gly Glu Gly Val Arg Gly Pro

30

Arg Arg Gln His Asn Pro Arg Arg Gln Gln Asp Pro Ser

50

Thr Leu Pro His Tyr Leu Gly Leu Gln Pro Asp Pro Asn

60

Gly Gly Gln Ile Gly Val Thr Ile Thr Ile Pro Leu Asn

70

Leu Gln Pro Pro Arg Val Leu Val Asn Leu Pro Gly Phe

90

80

Ile Thr Gly Pro Pro Leu Val Val Gln Gly Thr Thr Glu

100

14

Tyr Gln Tyr Gln Trp Gln Leu Thr Ala Pro Asp Pro Thr

110

Pro Leu Ser Asn Pro Pro Thr Gln Leu His Ser Thr Glu

120

130

Gln Ala Asn Thr Lys Thr Asp Ala Lys Ile Ser Asn Thr

140

Thr Ala Thr Thr Gln Asn Ser Thr Asp Ile Phe Glu Gly

Gly Gly Lys

and derivatives of this polypeptide.

10. Monoclonal and polyclonal antibodies directed against a peptide or polypeptide according to any one of the Claims 1 to 9.

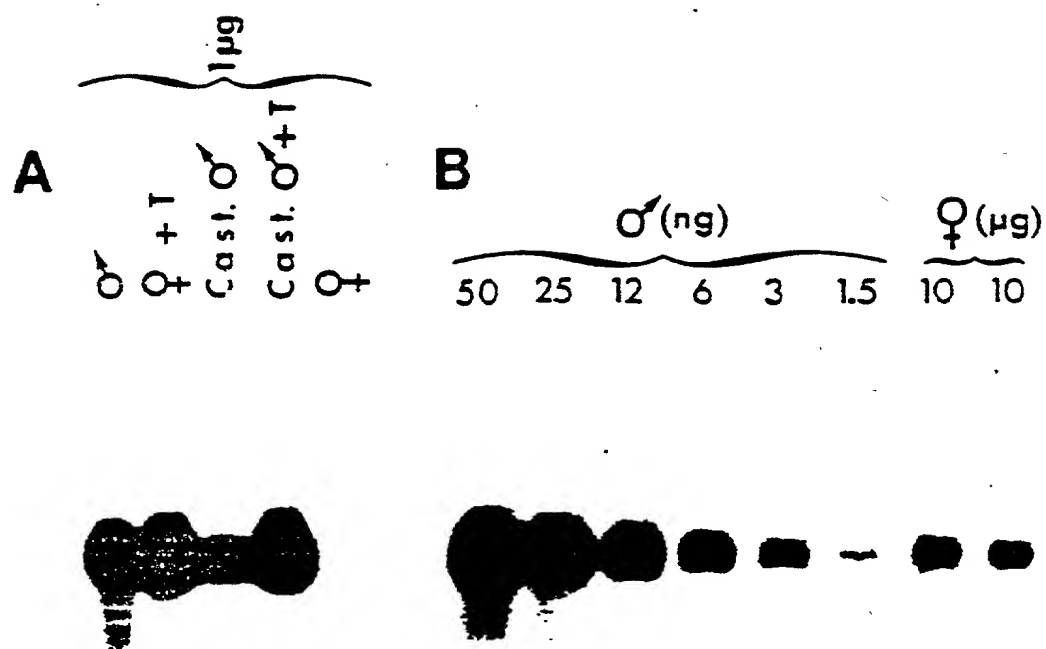
11. Hybridomas characterized in that they produce monoclonal antibodies according to Claim 10.

12. Assay or detection procedure for a peptide according to any one of the Claims 1 to 8, comprising the utilization of monoclonal antibodies directed against these peptides.

13. Assay procedure for a polypeptide according to Claim 9, comprising the utilization of monoclonal antibodies directed against this polypeptide.

14. Therapeutic composition containing a peptide or a polypeptide according to any one of the Claims 1 to 9.

1 / 1

FIG.1

SOLE/JOINT

DECLARATION AND POWER OF ATTORNEY

As a below named inventor, I hereby declare that my residence, post office address and citizenship are as stated below next to my name; that I verily believe I am the original, first and sole inventor (if only one name is listed below) or a joint inventor (if plural names are listed below) of the invention entitled: "PEPTIDES AND POLYPEPTIDES DERIVED FROM THE SUBMAXILLARY GLAND OF THE RAT, CORRESPONDING POLYCLONAL AND MONOCLONAL ANTIBODIES, CORRESPONDING HYBRIDOMAS AND USES OF THESE PRODUCTS FOR DIAGNOSIS, FOR DETECTION OR THERAPEUTIC PURPOSES"

☐ the attached application
(for original application)

☒ application Serial No. _____
filed 11 JUNE, 1990, and amended on _____

(for declaration not accompanying application)

that I have reviewed the above-identified application and understand the contents thereof, including the claims as amended by any amendment referred to above; that I acknowledge my duty to disclose information of which I am aware which is material to the examination of this application, i.e. such information where there is a substantial likelihood that a reasonable Examiner would consider it important in deciding whether to allow the application to issue as a patent; that I hereby claim foreign priority benefits under Title 35, United States Code §119 or §365 of any foreign application(s) for patent or inventor's certificate listed below and have also identified on said list any foreign application for patent or inventor's certificate on this invention having a filing date before that of the application on which priority is claimed:

Application Number	Country	Filing Date	Priority Claimed (yes NO)
88 13 353	FRANCE	88 13 353	

I hereby claim the benefit of Title 35, United States Code §120 of any United States application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in a listed prior United States application in the manner provided by the first paragraph of Title 35, United States Code, §112, I acknowledge my duty to disclose any material information of the type described above which occurred between the filing date of the prior application and the national or PCT international filing date of this application:

Application Serial No.	Filing Date	Status
PCT/FR89/00523	11 OCTOBER, 1989	(prior art) pending abandoned

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I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

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